

# Comparison of Direct and Indirect Dielectric Barrier Discharge Approaches for Upscaled Production of Plasma-Activated Water

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This study presents a comparative analysis of direct and indirect dielectric barrier discharge (DBD) approaches for the upscaled production of plasma-activated water (PAW). A cylindrical DBD reactor, consisting of nine high-voltage and nine ground electrodes operated with compressed air, was designed to investigate the generation of plasma and production of reactive oxygen and nitrogen species (RONS). The production of reactive species was evaluated through two distinct methods: indirect treatment, involving the dissolution of plasma-generated gas into water, and direct treatment, where the water flows through the active discharge region. Performance was assessed through the measurement of long-lived RONS (primarily hydrogen peroxide and nitrites), with results further analyzed through the determination of energy yields ( $g/kWh$ ) and pH. The results indicate that the direct treatment approach achieves significantly higher chemical production efficiency, identifying it as the more effective strategy for the upscaled production of PAW. The relative advantages and disadvantages of both direct and indirect approaches are discussed in the context of system optimization and industrial scalability.

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## I. Introduction

Plasma activated water (PAW) has emerged as an innovative technology with the potential to contribute significantly to sustainable development [1]. PAW is formed by treating water with an ionized gas, which activates water molecules to generate highly reactive species that can be used in a wide range of applications including medicine, water treatment, agriculture, wastewater treatment, and food processing [2, 3, 4, 5, 6]. One of the key advantages of PAW is that it is an environmentally-friendly alternative to conventional chemical treatments and processes. One such example is the Haber-Bosch process, which is used to produce synthetic fertilizers. This process involves high-energy consumption and results in significant greenhouse gas emissions and has a devastating impact on the environment [7, 8]. In contrast, PAW can be generated using green technology and poses less of a burden on the environment. It has immense potential to replace traditional methods of chemical treatments and promote the health and well-being of human beings.

The reason behind the significant interest in PAW in several applications is ascribed to the

production of numerous reactive oxygen and nitrogen species (RONS) such as hydrogen peroxide ( $H_2O_2$ ), nitrites ( $NO_2^-$ ), nitrates ( $NO_3^-$ ), nitric acid ( $HNO_3$ ), etc. These species are formed through the interaction of plasma components (such as electrons, ions, and UVs) with ambient gas and water molecules [2, 9, 10]. Furthermore, the chemistry of PAW including the pH can be specifically tailored by altering the feed gases and inclusion of reactive metals inside the liquid solution [11]. This makes PAW more attractive for use in several biomedical and agricultural applications.

The use of atmospheric pressure plasma sources for the production of PAW has been extensively investigated. However, the vast majority of these studies have been performed on very small liquid volumes (typically a few hundred microliters to a few milliliters) to explore the effects of operating parameters such as applied voltage, frequency and gas flow [2, 9, 12, 13]. While these laboratory-scale experiments have provided valuable mechanistic insights, they are inherently difficult to translate to industrial-scale throughputs required for practical applications in agriculture, wastewater treatment and food safety.

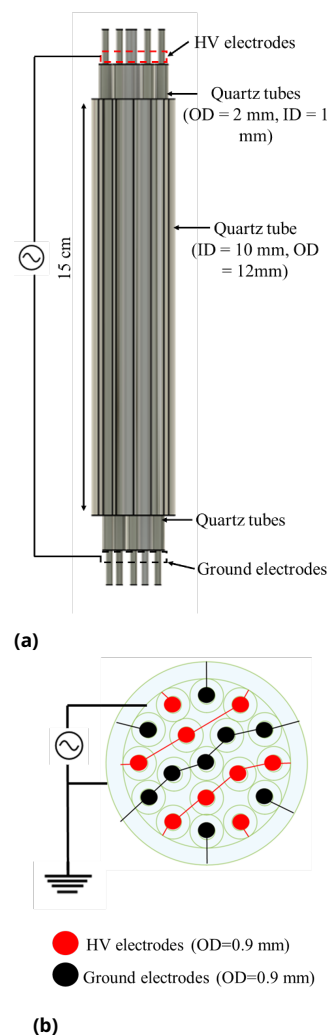
Although microbubble-enhanced cold plasma activation has been proposed as one promising route to improve gas-liquid mass transfer in flowing-water [14, 15, 16], this approach relies on complex Venturi injectors that can suffer from bubble coalescence, pressure drop, and long-term clogging, and still lacks systematic energy-yield benchmarking at larger scales. Critically, there remains a notable absence of comparative studies that directly evaluate direct (plasma in immediate contact with liquid) versus indirect (plasma-activated gas dissolved into liquid) treatment modes within the same upscaled reactor geometry. Such a head-to-head comparison is essential because direct interaction is expected to generate higher concentrations of reactive oxygen and nitrogen species (RONS) as a result of enhanced short-lived radical formation at the gas-liquid interface, while indirect treatment offers simpler operation but lower overall efficiency. Without this direct comparison, it is unclear which approach provides the optimal balance of RONS yield, energy efficiency, and operational simplicity for cost-effective, large-volume PAW production using inexpensive compressed air and portable power supplies.

In this article, we address this research gap by presenting a systematic comparison of direct and indirect dielectric barrier discharge (DBD) approaches for upscaled PAW production using a custom cylindrical reactor with eighteen stainless-steel electrodes.

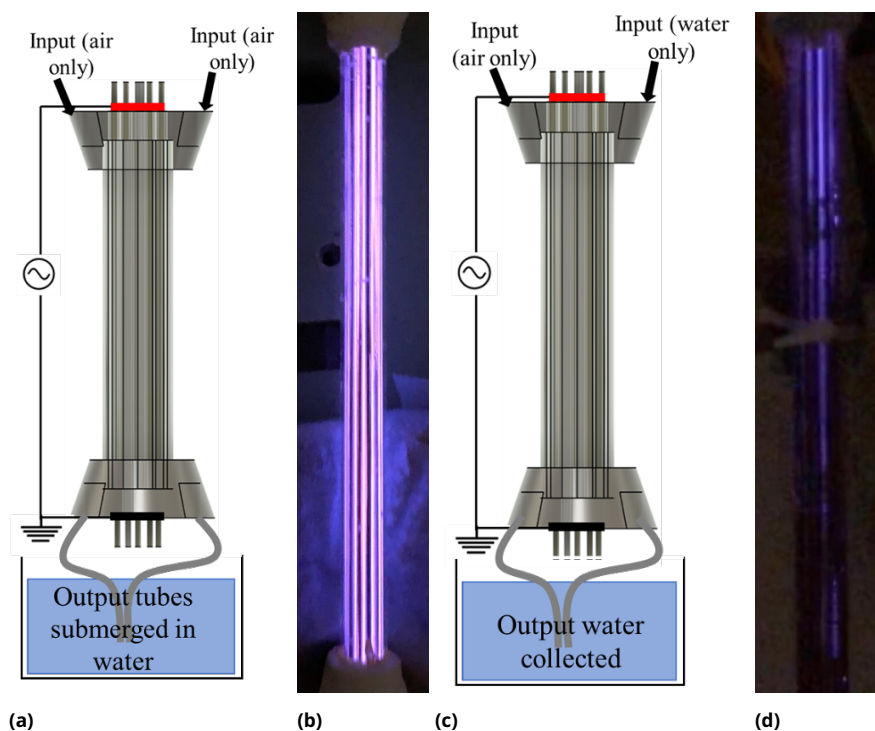
## II. Experimental Setup and Methodology

The schematic and the cross-sectional view of the reactor used for generating PAW is shown in Fig. 1a and 1b respectively. The cylindrical DBD reactor consists of eighteen stainless-steel wire electrodes (outer diameter, OD = 0.9 mm), each inserted inside individual quartz tubes (inner diameter, ID = 1 mm and OD = 2 mm). All these tubes are arranged inside a larger quartz tube (ID = 10 mm, OD = 12 mm) to generate plasma within a length of 15 cm. Nine quartz tubes containing stainless-steel wires are inserted from the top side of the quartz tube and the remaining nine quartz tubes are inserted from the bottom side following the schematic in Fig. 1b. About a 3 cm length of the stainless-steel wire remained outside of the smaller quartz tube in order to connect them to the high voltage (HV) and ground electrodes. The stainless-steel electrodes enclosed within quartz tubes remained physically isolated from both the

plasma-treated water and the gas-liquid interface. As quartz is chemically inert and acts as a dielectric barrier, direct contact between the electrodes and the acidic plasma-activated water is prevented, minimizing the possibility of metal ion contamination in the activated water. A 3D-printed ceramic cap was inserted on the top and bottom end of the larger quartz tube and sealed with Torr-seal epoxy to hold the smaller quartz tubes in place and to seal the wires inside their individual small quartz tubes to prevent direct contact between the water and the metal wire electrodes. The ceramic caps also had provisions for inserting two silicone tubes (OD = 6mm) for inputting the air or water inside the reactor through one end, and outputting the plasma activated air or water through the other end. Generation of plasma was done by applying sinusoidal voltage using a commercial power supply (PVM-500, Information Unlimited, USA) between



**Figure 1:** Schematic of: (a) the DBD reactor used to generate PAW; and (b) the arrangement of eighteen electrodes within the DBD reactor.



**Figure 2:** Methods of generating PAW through: (a) indirect (plasma-activated gas dissolved into liquid), and (c) direct (plasma in immediate contact with liquid) methods. Photographs of the discharge during operation for: (b) indirect, and (d) direct modes are also shown.

the HV and the ground electrodes. The liquid used was distilled water (conductivity:  $\approx 3 \mu S.cm^{-1}$ ).

The formation of RONS in PAW was investigated using two methods. In the first method (Fig. 2a), compressed air at a flow rate of 20 standard liters per minute (SLPM) was passed into the reactor through the two inlet tubes. The applied voltage was fixed at  $\approx 10 kV$  at  $\approx 33 kHz$ . Under these conditions, a uniform glow discharge was formed between the HV and the ground electrodes to generate RONS from the air (Fig. 2b). The outlet tubes on the bottom side of the reactor were submerged inside a conical tube that contained fixed volume (25 mL) of water. In this configuration, RONS formed in the discharge region were carried by the gas flow and dissolved inside water, thus activating it indirectly. The indirect activation of water within a fixed volume was conducted for 5, 10, 20 and 30 minutes, and the characteristics of PAW were analyzed. In the second method (Fig. 2c), one inlet tube carried compressed air (flow rate: 50 SLPM), and the other inlet carried water into the reactor. The flow of water inside the reactor was continuous (flow rate:  $\approx 25$  mL/min). As water was also flowing inside the reactor, the higher flow rate of air created water bubbles inside the reactor, thus facilitating the formation of plasma. However, the plasma formed with the air and water flow from the

inlet tubes Fig. 2d was very different from the one with only air flow as seen in Fig. 2b. Capturing the plasma inside individual bubbles was challenging because the discharge points continuously shift as the bubbles move and evolve. The discharge forms bright filaments or glow regions within the bubbles, which are transient and dynamically distributed during operation. In this configuration, water was directly activated within the reactor, and the activated water was collected from the outlet tubes at the bottom of the reactor. The direct activation of water with continuous flow was also conducted for 5, 10, 20 and 30 minutes, and characteristics of PAW were analyzed.

Different gas flow rates were used for the indirect and direct treatment configurations due to operational constraints related to plasma stability and species collection. In the indirect treatment mode, a flow rate of 20 SLPM was used to allow sufficient residence time and efficient dissolution of plasma-generated reactive species into water during the collection process. In the direct treatment mode, where water flows through the reactor, a higher gas flow rate of 50 SLPM was required to sustain a stable gas-phase discharge and prevent plasma quenching by the liquid medium. These flow rates represent the operational conditions necessary for stable discharge and reliable data

collection within the current reactor geometry. To ensure a meaningful comparison between the two treatment modes, the measured concentrations of reactive species were normalized as described in the Results section.

Electrical signals of the discharge were acquired by connecting a Tektronix voltage probe (Model no. P6015) to the HV tungsten electrode and a Pearson current probe (Model no. 2100) to the ground electrode. The signals from the voltage and current probes were recorded in an oscilloscope (Tektronix MDO3024). Optical emission spectroscopy of the discharge was conducted using a 0.5-m Princeton Instruments' SP2500 spectrometer coupled to a PI-MAX 4 1024×256 pixel intensified charged coupled device (ICCD) camera. The spectrometer was calibrated in wavelength and intensity using the manufacturer provided calibration lamps (Intellical, Princeton Instruments).

The chemistry of PAW after direct and indirect plasma treatment was analyzed by measuring the concentration of long-lived RONS ( $\text{H}_2\text{O}_2$  and  $\text{NO}_2^-$ ) and pH. For  $\text{H}_2\text{O}_2$  measurements, a calibration curve with known concentrations of  $\text{H}_2\text{O}_2$  was constructed (see Fig. A1 in the appendix) by using a colorimetric assay based on o-phenylenediamine (OPD, CAS number: 95-54-5, Sigma Aldrich Corporation) and horseradish peroxidase (HRP, CAS number: 9003-99-0, Sigma Aldrich Corporation). HRP catalyses the oxidation of OPD in the presence

of  $\text{H}_2\text{O}_2$  to form 2,3-diaminophenazine which has an absorbance maximum at 450 nm (measured using SpectraMax iD5, Molecular Devices). The line of best fit obtained from the calibration curve was then used to calculate the concentration of  $\text{H}_2\text{O}_2$  in PAW [12, 13].

Measurement of  $\text{NO}_2^-$  in PAW also involved constructing a calibration curve with known concentrations of nitrite with the help of a commercially available kit [12]. We used a Griess reagent kit (Supplier: Sigma Aldrich, EC number: 215-981-2) that gives an absorbance maximum of 540 nm (measured using SpectraMax iD5, Molecular Devices) upon reaction with the nitrites. The calibration curve constructed with known concentrations of  $\text{NaNO}_2$  is shown in Fig. A2 (appendix). The line of best fit obtained from the calibration curve was then used to calculate the concentration of  $\text{NO}_2^-$  in PAW formed through indirect and direct plasma treatments.

Measurement of pH involved immersing a pH indicator paper (catalogue number: 110962, Merck Millipore) in PAW (Volume = 100  $\mu\text{l}$ ) and referencing the change in color, as seen with the naked eye, to the corresponding pH value on the chart provided in the kit.

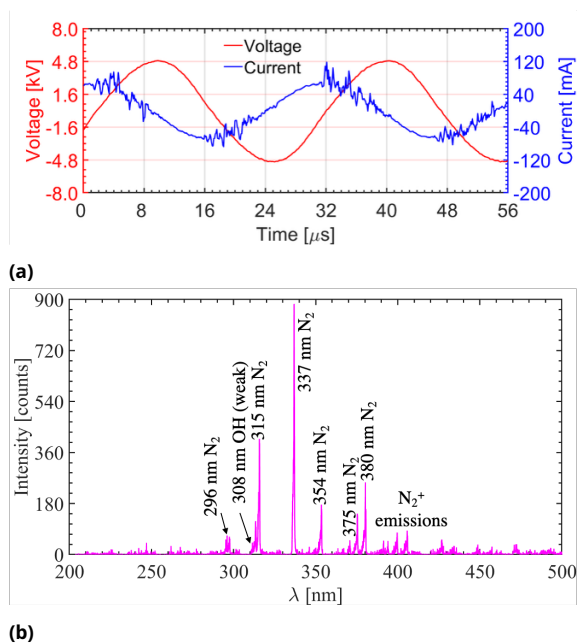
### III. Results and Discussion

Fig. 3 shows the electrical and optical emission characteristics of the discharge. Operation of the cylindrical DBD is done using a sinusoidal power supply with an applied voltage of  $\approx 10$  kV (p-p), rms current  $\approx 35$  mA, frequency  $\approx 33$  kHz. The current (I) and voltage (V) waveforms of the discharge for the cylindrical DBD operating with air flow is shown in Fig. 3a. The IV-waveforms over one time period (T) were used to calculate the dissipated power (P) as:

$$P = \frac{1}{T} \int_{t=0}^{t=T} I(t)V(t)dt \quad (1)$$

The dissipated power calculated using Eqn. (1) was 18.26 W and 22.63 W for the indirect and direct treatment methods, respectively.

Optical emission spectrum of the discharge recorded using a 0.5-m Princeton Instruments' SP2500 spectrometer coupled to a PI-MAX 4 1024×256 pixel intensified charged coupled device (ICCD) camera is shown in Fig. 3b. The spectra were recorded during indirect preparation of PAW (Fig. 2a). Several bands of nitrogen second positive system ( $\text{N}_2$  SPS) formed through the dissociation of molecular nitrogen at 296 nm, 315 nm, 337 nm,



**Figure 3:** (a) Electrical and (b) Optical emission characteristics of the discharge recorded during indirect treatment of water with the cylindrical DBD.

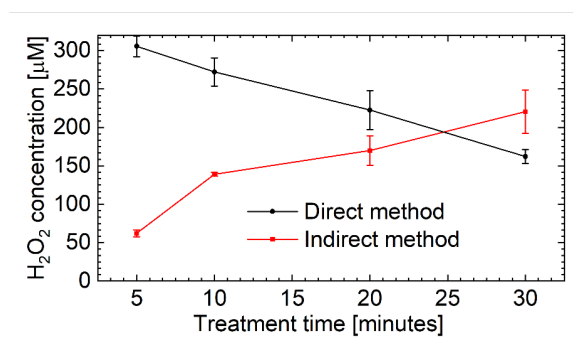
354 nm, 375 nm, 380 nm, 394 nm, etc. are clearly observed [17, 18, 19, 20]. Emissions from nitrogen first negative system ( $N_2$  FNS) are also observed around 391 nm. In addition to excited nitrogen species, there is also weak emission from hydroxyl radical ( $\bullet OH$ ) at 308 nm [21, 22]. Excited nitrogen and  $\bullet OH$  are primary species which are responsible for the formation of long-lived RONS in PAW. The measurement of hydrogen peroxide ( $H_2O_2$ ) and nitrites ( $NO_2^-$ ) will be described in the next paragraphs. While OES was not recorded during direct plasma treatment due to temporal and spatial instability caused by dynamic bubble formation, the discharge characteristics in this mode are expected to differ from those observed in the gas-phase configuration. In the direct treatment mode, plasma is generated within gas bubbles surrounded by liquid water, where interactions with a moisture-saturated environment and the gas-liquid interface may enhance the formation of hydroxyl radicals and other hydrogen-related reactive species compared to the indirect mode [23].

For the indirect method, the emission from  $N_2$  SPS between 370 - 380 nm was also used to estimate the rotational and vibrational temperature of the DBD discharge using MASSIVEOES [24, 25, 26]. At atmospheric pressure, because of the high collision frequency, the gas temperature of the plasmas is close to rotational temperature. The rotational and vibrational temperature of the discharge were found to be  $\approx 433$  K and 3422 K respectively (Fig. A3, appendix). We did not record the OES of the discharge during direct plasma treatment of the flowing water (Fig. 2b), as the regions of plasma formation changed frequently with time due to bubble formation. But, due to the continuous flow of the water, the temperature of

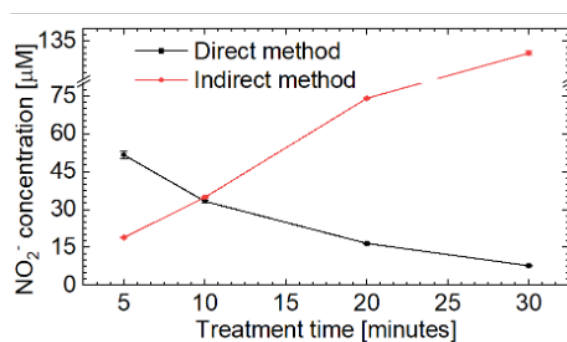
the external quartz tube was at room temperature and was monitored with a thermocouple. The temperature of the PAW collected through direct plasma treatment also remained at room temperature.

Fig. 4 shows the results for  $H_2O_2$  concentrations in PAW. In the indirect method, the concentration of  $H_2O_2$  increased with the treatment time up to 30 minutes. After 5 minutes, the concentration of  $H_2O_2$  is  $\approx 62 \mu M$ . This increased to  $\approx 220 \mu M$  when the plasma exposure time was increased to 30 minutes. On the other hand, the concentration of  $H_2O_2$  formed through direct method kept on decreasing. At 5 minutes,  $\approx 305 \mu M$  of  $H_2O_2$  was formed in PAW. This decreased by almost half ( $\approx 160 \mu M$ ) when the plasma exposure time was increased to 30 minutes.

The measurements of  $NO_2^-$  concentrations for the indirect and direct methods inside PAW are shown in Fig. 5. Although, there are several nitrogen derivatives such as nitrates, nitric acid, peroxyntrous acid, etc. formed inside PAW, we measured  $NO_2^-$  to get an overall indication of reactive nitrogen species formed inside PAW. At pH values close to neutral,  $NO_2^-$  is known to be a dominant detectable species before full disproportionation into  $NO_3^-$  [27]. The concentrations of  $NO_2^-$  measured for indirectly and directly treated water are shown in Fig. 5. The trend in the  $NO_2^-$  concentrations are similar to those of  $H_2O_2$  concentrations for both methods. For indirect method, the concentration of  $NO_2^-$  increased from  $\approx 19 \mu M$  to  $\approx 130 \mu M$  when the plasma exposure time was increased from 5 minutes to 30 minutes respectively. For the direct method, these concentrations decreased from  $\approx 51 \mu M$  to  $\approx 8 \mu M$  when the plasma treatment time increased from 5 minutes



**Figure 4:** Concentrations of  $H_2O_2$  measured at plasma exposure time of 5, 10, 20 and 30 minutes with the direct and indirect methods. The calibration curve used for this measurement is in the appendix (Fig. A1)



**Figure 5:** Concentrations of  $NO_2^-$  measured at plasma exposure time of 5, 10, 20 and 30 minutes with the direct and indirect methods. The calibration curve used for this measurement is in the appendix (Fig. A2)

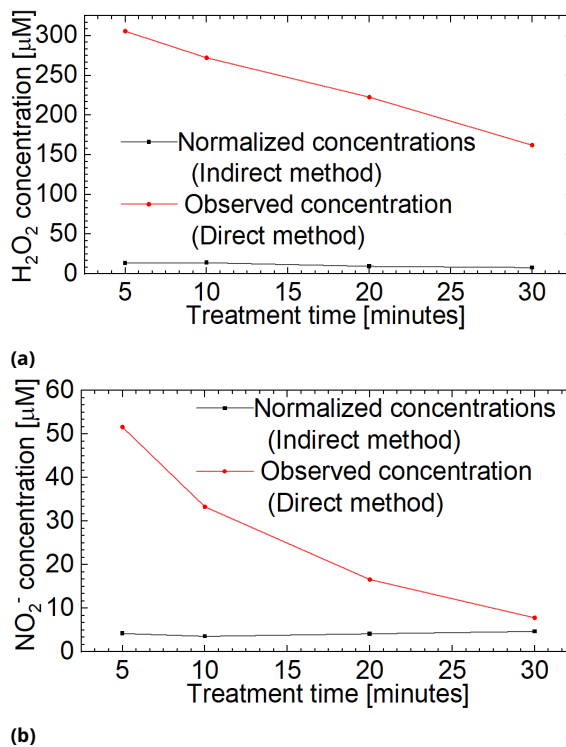
to 30 minutes respectively.

The decrease in the concentrations of  $H_2O_2$  and  $NO_2^-$  for water activated through direct method at different treatment times in Fig. 4, 5 are very interesting. It should be noted that the volume of the plasma activated water coming from the outlet tubes at different treatment times increased with time as the water was constantly pumped inside the reactor by the motor through the inlet tubes. After 5 minutes,  $\approx 113$  mL was collected in the container. This increased to  $\approx 700$  mL after 30 minutes of plasma treatment. During water flow, the formation of plasma inside the reactor only takes place inside the air bubbles. The plasma in this case is not formed at all points (as also depicted from the photograph in Fig. 2d). The formation of plasma at only few positions inside the reactor suggests that all water passing inside the reactor is not activated by the DBD plasma. We speculate that the volume of water not activated by plasma increased with time and this led to the decrease in the concentrations of  $H_2O_2$  and  $NO_2^-$ . On the other hand, for the indirect treatment - the volume of water in the container was 25 mL and this became more concentrated when the outlet tubes were continuously submerged inside the water. This led to the higher concentration of both  $H_2O_2$  and  $NO_2^-$  in the plasma activated water.

To further understand the efficacy of RONS production inside PAW by the indirect and direct methods, we normalized the concentrations of  $H_2O_2$  and  $NO_2^-$  obtained through indirect method with the total volume of water that was collected through direct method. The normalized concentration (N) for indirect plasma treatment were calculated to account for the concentration of  $H_2O_2$  and  $NO_2^-$  that could have been formed in a total volume of collected water using direct treatment  $V_1$  using known parameters (concentration ( $C_1$ ) and volume ( $V_1$ )) for the indirect plasma treatment. This was calculated using eqn. (2) as:

$$N[\mu m] = C_1[\mu m] \times \frac{V_1[mL]}{V_2[mL]} \quad (2)$$

This normalization assumes that the total mass of RONS generated by the plasma under similar discharge conditions is conserved and that the observed concentration difference arises primarily from dilution in different liquid volumes. In practice, gas-liquid mass transfer and dissolution efficiency may also influence the measured concentrations, and therefore the normalized values should be interpreted as an approximate comparison between the two treatment modes. The results of normalization are shown in Fig. 6a,



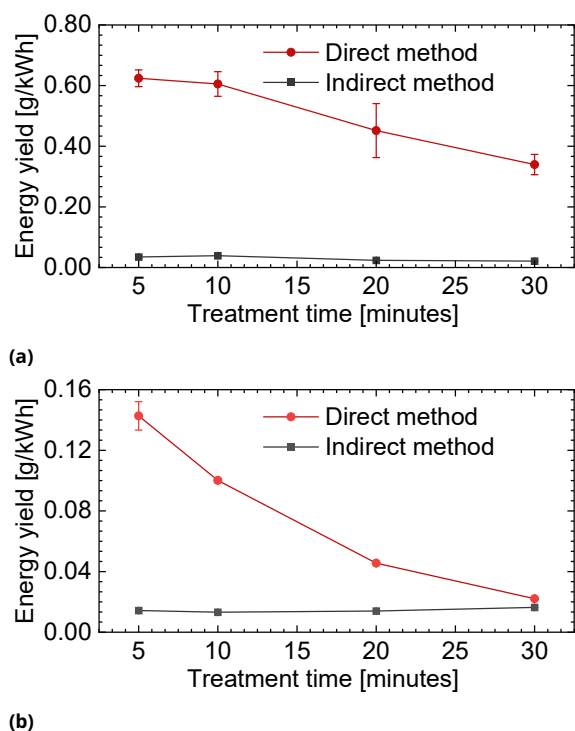
**Figure 6:** Normalized and observed concentrations of (a)  $H_2O_2$  and (b)  $NO_2^-$  from indirectly and directly treated water at different plasma treatment times. The normalized values represent the concentration of  $H_2O_2$  and  $NO_2^-$  that could have been formed when the volume of the treated water with the indirect DBD was the same as the volume of activated water collected through direct plasma treatment.

6b, and also summarized in the table (see table 1, 2 in the Appendix). Although the concentrations of both  $H_2O_2$  and  $NO_2^-$  decreased with plasma exposure time during the direct plasma treatment, these were much higher than the normalized concentrations that could be obtained using indirect method in the same volume of target liquid. This suggests that direct treatment of water with the cylindrical DBD could be a better choice for making PAW.

We also evaluated the efficiency of direct and indirect methods by calculating the energy yield of  $H_2O_2$  and  $NO_2^-$ . This energy yield represents the amount of chemical species generated per unit electrical energy supplied to the plasma reactor. The energy yield ( $EY$ ) was calculated as:

$$EY[g/kWh] = \frac{C[mol/L].V[L].M[g/mol]}{P[kW].t[h]} \quad (3)$$

where  $C$  is the measured molar concentration in  $mol/L$ ,  $V$  is the volume of treated water in  $L$ ,  $M$  is the molar mass of the species in  $g/mol$ ,  $P$  is

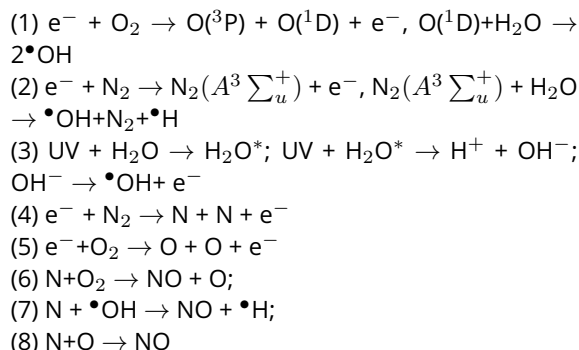


**Figure 7:** Estimation of energy yield for (a) H<sub>2</sub>O<sub>2</sub> and (b) NO<sub>2</sub><sup>-</sup> from indirect and direct treatment approaches at different plasma treatment times.

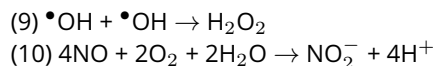
the dissipated plasma power in  $kW$ , and  $t$  is the treatment time in  $h$ . The results for 5-30 minutes of plasma treatment are presented in Fig. 7. Overall, the energy efficiency was heavily influenced by the treatment approach. The direct treatment approach [despite decrease in EY from  $0.62g/kWh$  ( $t=5$  minutes) to  $0.34g/kWh$  ( $t=30$  minutes)] yielded H<sub>2</sub>O<sub>2</sub> concentrations approximately 16-20 times higher than those achieved via the indirect approach (Fig. 7a), identifying the direct approach as a superior strategy for the energy-efficient production of hydrogen peroxide. This enhanced efficiency is likely due to the immediate contact between the plasma and the water molecules. On the other hand, a distinct time-dependent decay in EY was observed for NO<sub>2</sub><sup>-</sup> production though direct treatment approach was more efficient than the indirect one (Fig. 7b). Specifically, the EY for NO<sub>2</sub><sup>-</sup> under direct treatment was ca. 10 times higher than the indirect method at  $t=5$  minutes; however, this factor decreased to 7.60, 3.26 and 1.34 at 10, 20 and 30 minutes, respectively. The diminishing EY over time suggests that as the concentration of reactive species increase with time, secondary loss mechanisms could become more dominant.

The reason for higher concentrations of both H<sub>2</sub>O<sub>2</sub> and NO<sub>2</sub><sup>-</sup> in the directly activated water with the cylindrical DBD are due to the differences in

the mode of activation. Formation of H<sub>2</sub>O<sub>2</sub> and NO<sub>2</sub><sup>-</sup> inside the DBD reactor mainly takes place through the dissociation of H<sub>2</sub>O, O<sub>2</sub> or N<sub>2</sub> below [28, 2, 29, 19, 30, 12]:

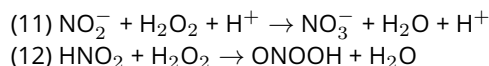


The  $\bullet OH$  and NO so formed in (1)-(8) recombine to form H<sub>2</sub>O<sub>2</sub> and NO<sub>2</sub><sup>-</sup> as follows [31, 32]:



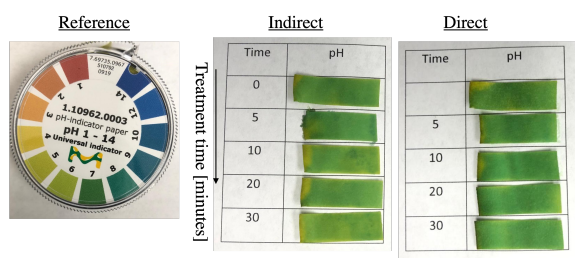
During direct plasma treatment, most of the reactions [(1)-(10)] take place within the DBD reactor where high energy electrons and excited species interact directly with the ambient gas species (H<sub>2</sub>O, N<sub>2</sub>, O<sub>2</sub>). As a result, H<sub>2</sub>O<sub>2</sub> and NO<sub>2</sub><sup>-</sup> could be formed instantaneously within the discharge region. On the other hand, during indirect plasma treatment, it is mainly the plasma activated gas that interacts with the target liquid to result in the production of H<sub>2</sub>O<sub>2</sub> and NO<sub>2</sub><sup>-</sup>. Since the lifetime of short-lived reactive species (such as  $\bullet OH$ , O, N, etc.) is short [32, 28], only the long-lived species formed in the discharge region are carried/dissolved onto the target water resulting in the lower concentration of both H<sub>2</sub>O<sub>2</sub> and NO<sub>2</sub><sup>-</sup>.

NO<sub>2</sub><sup>-</sup> or HNO<sub>2</sub> formed in PAW can also combine with H<sub>2</sub>O<sub>2</sub> to form NO<sub>3</sub><sup>-</sup> or ONOOH as [12]:



Although, the concentration of NO<sub>3</sub><sup>-</sup> was not explicitly measured in this work, the simultaneous decrease in EY of both H<sub>2</sub>O<sub>2</sub> and NO<sub>2</sub><sup>-</sup> (Fig. 7) suggests that reactions (11)-(12) could be responsible for the secondary loss.

The final set experiments were designed to investigate the effect of plasma treated solutions (indirect and direct) on pH. pH is an important parameter that affects the growth of plants [33]. The level of pH plants require for growth may be different but most of the plants prefer to grow



**Figure 8:** pH of PAW prepared through indirect and direct treatment of water.

around neutral pH. The pH of PAW normally goes to acidic level upon exposure to plasma treatment. For example, the pH of 5 minutes PAW prepared with plasma jet is between 2-3 [12]. In this work, a pH indicator paper was primarily used to provide a visual confirmation of the pH change in plasma-activated water. Although this method provides only semi-quantitative estimation, it effectively illustrates the relative pH difference between untreated and plasma-treated samples. Despite activating for up to 30 minutes, the pH of PAW prepared at different treatment times with both indirect and direct methods remained close to neutral (Fig. 8) and this could favor the growth of several plants.

#### IV. Conclusions

In conclusion, a comparison on the characteristics of PAW prepared through indirect and direct activation of water by a cylindrical atmospheric pressure DBD reactor operated with compressed air is presented in this work. The DBD operated with only air flow runs at higher gas temperature leading to heating of the DBD reactor. On the other hand, the flow of liquid inside the DBD reactor results in the lower heating of the reactor but formation of plasma is not uniform. Results obtained from the chemical analysis of PAW suggest that concentration of  $H_2O_2$ ,  $NO_2^-$  (and other RONS that could be present inside PAW) are higher from directly activated PAW which were verified through the normalization experiments and calculation of energy efficiencies. The pH of the PAW prepared using both methods remained close to neutral. The approach presented in this study could be scaled towards production of PAW in large quantities for applications in agriculture and medicine.

#### V. Acknowledgement

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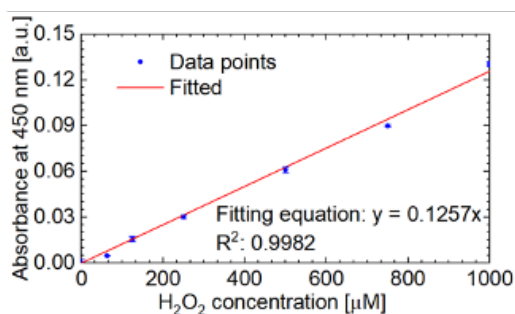
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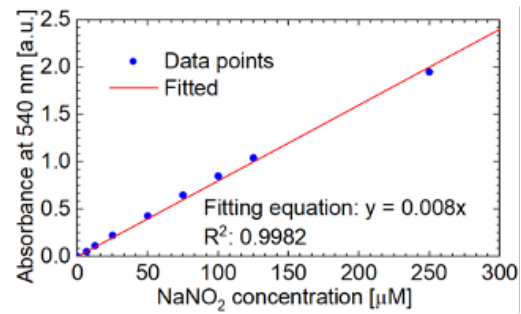
## VII. Data Availability Statement

All data that support the findings of this study are included within the article and its appendix.

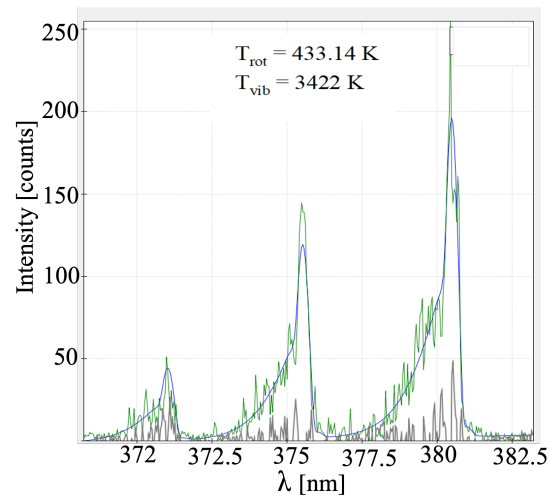
## VIII. Appendix



**Figure A1:** Calibration curve constructed with known concentrations of H<sub>2</sub>O<sub>2</sub> for estimating the concentration of H<sub>2</sub>O<sub>2</sub> in PAW.



**Figure A2:** Calibration curve constructed with known concentrations of NaNO<sub>2</sub> for estimating the concentration of NO<sub>2</sub><sup>-</sup> in PAW.



**Figure A3:** Measurement of rotational and vibrational temperature of the DBD discharge (for indirectly preparing PAW) using MASSIVEOES.

T [min]	C <sub>1</sub> [μM]	V <sub>1</sub> [mL]	V <sub>2</sub> [mL]	N [μM]
5	62.22	25	113.33	13.72
10	139.12	25	246.67	14.10
20	169.87	25	450.00	9.44
30	220.53	25	696.67	7.91

**Table 1:** Normalized concentrations of H<sub>2</sub>O<sub>2</sub> for the indirect treatment by considering the volume of liquid collected with direct plasma treatment. The normalized values represent the concentration of H<sub>2</sub>O<sub>2</sub> and NO<sub>2</sub><sup>-</sup> that could have been formed when the volume of the treated water with the indirect DBD was the same as the volume of activated water collected through direct plasma treatment.

T [min]	$C_1$ [ $\mu\text{M}$ ]	$V_1$ [mL]	$V_2$ [mL]	N [ $\mu\text{M}$ ]
5	18.94	25	113.33	4.18
10	34.90	25	246.67	3.54
20	74.10	25	450.00	4.12
30	130.24	25	696.67	4.67

**Table 2:** Normalized concentrations of  $\text{NO}_2^-$  for the indirect treatment by considering the volume of liquid collected with direct plasma treatment. The normalized values represent the concentration of  $\text{H}_2\text{O}_2$  and  $\text{NO}_2^-$  that could have been formed when the volume of the treated water with the indirect DBD was the same as the volume of activated water collected through direct plasma treatment.



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